

Design and performance of unique field chamber and CO₂ control system for the investigation of biological impacts on wheats grown under fluctuating CO₂

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Abstract

The CO₂ concentration [CO₂] in the atmosphere is increasing, affecting plant growth and development. Free Air CO₂ Enrichment (FACE) technology is used for the study of plant development under elevated CO₂. However, criticism of the FACE technology is that it underestimates plant and crop responses to elevated [CO₂] because rapidly fluctuating [CO₂] within the FACE rings may force stomata to close more often and for longer periods than would occur under non-FACE conditions. Therefore, data from FACE rings cannot be used as validation for plant and crop responses in crop modelling. This report focuses on the engineering of chamber design to test constant vs. variable changes in [CO₂] levels on crops. Three chambers were built, and a non-replicated study was conducted during the 2015 and 2016 seasons to assess the methodology and chamber design. The chambers were set up to test the impacts of constant ambient [CO₂], elevated [CO₂] and variable [CO₂] on wheat biomass, yield and water use efficiency. The chamber design and control system performed as expected keeping 1 min average CO₂ concentration within 1.8 μmol/mol (insignificant variation) to the set point of 610 μmol/mol for elevated CO₂ treatments (fluctuating and constant).

Key Words

FACE System, fluctuating CO₂, field chamber, biological impact, wheat, Australia

Introduction

Naturally occurring CO₂ concentration [CO₂] in the atmosphere is increasing and currently is around 410 μmol/mol (Mauna Loa Observatory 2019). Elevated CO₂ (e[CO₂]) affects plant growth and development in various ways, most notably by increasing biomass, yields and water use efficiency (Leakey *et al* 2009, Taub 2010). Rapidly fluctuating [CO₂] within the FACE rings may force stomata to close more often and for longer periods than would occur under non-FACE conditions (Cardon *et al* 1994, Holtum and Winter 2003, Bunce 2012) leading to underestimates of plant and crop responses to elevated [CO₂] (Tubiello and Ewert 2002, Tubiello *et al* 2007, Ainsworth *et al* 2008). There have been a few studies (Hendry *et al* 1997, Holtum and Winter 2003, Bunce 2012) investigating whether high frequency fluctuations of [CO₂] affect plant response. Results varied between research teams.

To understand better the biological impact of high frequency fluctuations of [CO₂] on wheat, custom chambers were designed and built to mimic (as closely as possible) the fluctuations found inside the rings of the Australian Grains Free Air CO₂ Enrichment (AGFACE) facility. Baker *et al* (2009, 2014a, 2014b) field tested open system chambers and reported that the chambers were able to accurately estimate canopy transpiration for many field applications such as comparison of canopy gas exchanges and water use efficiencies amongst irrigation treatments. Consequently, in 2015, three chambers (Figure 1) were constructed based on the design theme of Baker *et al* (2014a & 2014b) and an experiment conducted for two seasons. Three treatments were included: 1) ambient [CO₂], 2) elevated constant [CO₂], and 3) elevated fluctuating [CO₂]. The details of chamber design and performance are presented in this article.

Methods

Chamber design

The chamber frames were made of 20 mm x 20 mm x 1.6 mm RHS welded in shape. The volume of the main chamber was 1 cubic metre with two inlet and outlet funnels connected (Figure 1). The whole chamber was clad with 2 mm thick solid clear UV resistant polycarbonate sheets (Laserlite Australia Pty Ltd Victoria Australia). A portable 300 mm diameter ventilator (MPV 300, Fanmaster, Sydney, Australia) with airflow capacity of 1300 L/s was used to suck the air through the chamber at a speed of about 1.3 m/s. A purpose-built cover protected the ventilator's electric wiring from harsh outdoor weather, especially the rain. This high volume, low pressure fan was fitted to each chamber to maintain the average air temperature inside the chambers within ±1 °C to ambient. Two clear 2 mm thick polycarbonate barriers were used at both inlet and outlet sides of the chamber to control the air flow and air distribution through the chamber. There was a 90

mm gap at the bottom of the barrier at the inlet side and a 90 mm gap at the top of the barrier at the outlet side. This arrangement was found to be the best for the desired airflow and air distribution after trying several ideas, including perforated clear plastic sheets with different sizes and orientations of holes. Spouts were made by a local engineering workshop to attach 300 mm diameter flexible ducting at the inlet side and the ventilator fan at the outlet side. The inlet duct was raised 2 m above the ground to minimise contamination from expelled CO₂ gas at the outlet side (Figure 1).

CO₂ Sensors

Chambers 1 & 2: The [CO₂] in chambers 1 & 2 were monitored by factory-calibrated infrared gas analysers (SBA-4model, PP Systems, USA) with the sampling head placed inside the chambers (Mollah *et al.* 2009). **Chamber 3:** Open Path LI - 7500A CO₂/H₂O Gas analyser (LI-COR®, Inc, Lincoln, Nebraska 68504 USA) was used to monitor the fluctuating [CO₂] in chamber 3 (Figure 1). The analyser was set to sample 5 times per second. The high speed was needed so that the fluctuating CO₂ could be controlled quickly.

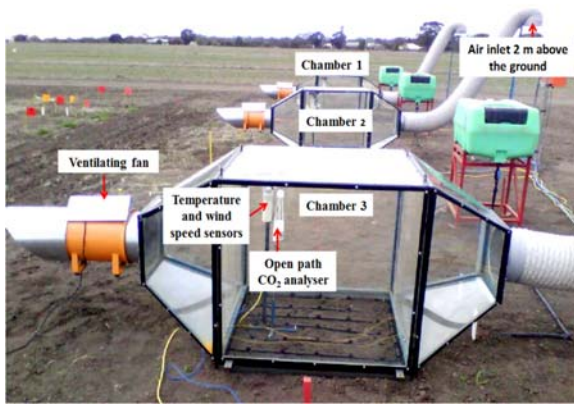


Figure 1. Chambers to study biological impacts of wheat grown under fluctuating CO₂.

CO₂ control system

To compare the biological impact of high frequency fluctuations of [CO₂] on plants, the criteria set out for the chambers in the field were: 1) No CO₂ injection and its control needed for Chamber 1 (ambient, (~400 μmol/mol)), b) CO₂ injection and the control of CO₂ levels needed to maintain a set constant [CO₂] at 610 μmol/mol in Chamber 2, c) CO₂ injection and the control of CO₂ levels needed to maintain fluctuating [CO₂] between 400 – 1200 μmol/mol with a median value of 610 μmol/mol and 30 s periods. The architecture of the CO₂ control system consisted of the assembly of sensors, modems, computers etc. (Figures 2 & 3). The [CO₂], wind speed, %RH and temperature from each chamber and ambient temperature were received by the data hub every second (Figure 2) and then sent to a field office computer for logging (Figure 3). An alarm system was set up to send an email to a nominated account (configurable) if power failed and or wind-speed in any chamber dropped below a set limit.

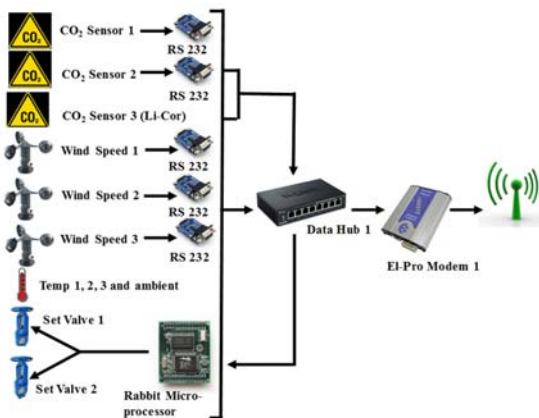


Figure 2. Schematic diagram of the components of the control system in the field.



Figure 3. Schematic diagram of the equipment architecture in the field office.

Rabbit Firmware and control valves

Rabbit firmware was used in Chambers 1 & 2 and consisted of: CPU SR9150, Backplane chassis SR9010, A/D converter. 0 - 10 V SR9300, Relay - 6 SR9500, LCD Keyboard/Display SR9010 +SR9050, FWT 18W SR9300 and Timer/Counter Card SSCTC-LS. The Rabbit Microprocessor (written in dynamic C) received [CO₂] from chambers 2 & 3 to operate the valves which were controlling the injection of CO₂ to chamber 2 & 3 as per the program. Two SMC ITV1030-011S5 valves with input 4~20mA DC, output 5 - 500 kPa were

used for the control of the [CO₂] in Chamber 2 & 3. One SMC VT307 - 5D -02, 0 - 0.9 MPa valve was used to shut down the CO₂ supply at night and turn on in the morning.

Understanding the CO₂ fluctuation in existing FACE Rings

To determine the target [CO₂] for the chambers, an open-path CO₂ analyser (Li-COR used in chamber 3) was placed half way between the edge and the ring-centre in an existing 12 m FACE ring to record CO₂ concentration @ 20 readings per second over a few days. A representative 5s recoding provided the basic understanding of wide CO₂ fluctuation inside an AGFACE ring. About 98% fluctuation happened between 400 and 1200 μmol/mol with a median CO₂ concentration value of 610 μmol/mol. This value was used as the set value for chamber 2 (constant CO₂) and the median value for the fluctuation profile (between 400 and 1200 μmol/mol) of chamber 3.

Data Logging and downloading

The data logging strategies were: a) Raw data (as observed) - one file per day, b) Raw data (as observed) – one file per hour, c) 1-second average data files - one file per day, d) 1-minute average data files - one file per day, e) 1-minute average cumulative data file - one file for whole season (append data to a file at midnight). The PC at the site office was accessed remotely via LogMeIn program for data downloading.

Monitoring system performance

System performance was regularly monitored using a dashboard (written in LabView) on computer screen (Figure 4) and alarms received on mobile phone. There was a provision to access the dashboard remotely to facilitate monitoring from a PC away from the field site. The dashboard also had a specific button to monitor the performance of chamber 3, where CO₂ was set to fluctuate within a given range. This was very useful as the set up for CO₂ was affected by the growth of the canopy inside the chamber (Figure 5

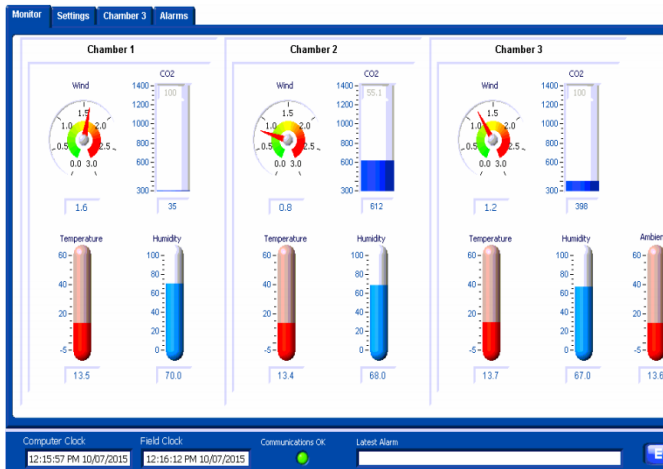


Figure 4. Dashboard on computer screen to monitor system performance and trouble shooting.

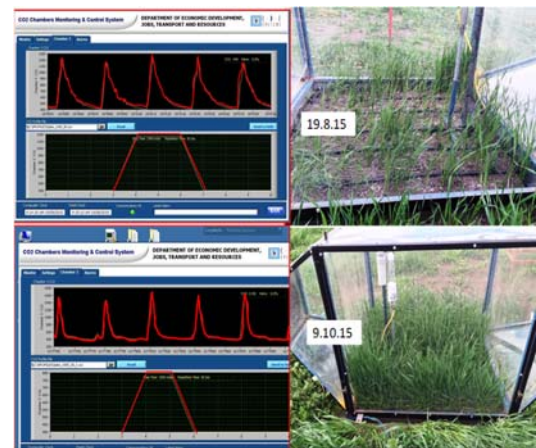


Figure 5. Chamber 3 performance at different growth stage

Results

The system was able to maintain 1-min average [CO₂] very close to the set point of 610 μmol/mol for chambers 2 (constant, 611.8 μmol/mol) & 3 (fluctuating, 610.6 μmol/mol) [Figure 6]. There were no significant differences between the chambers for temperature and relative humidity (RH) and as expected humidity reduced with increasing temperature (Figure 7). The temperatures and RH (Figure 7) in the chambers closely followed the ambient temperature and ambient RH (results not shown). These results suggest that the ventilator chosen to suck the air through the chamber at a speed of about 1.3 m/s was sufficient and kept the chamber temperature and RH close to the ambient. The control of CO₂ levels in the chambers (outlined in Methods) worked as expected. The CO₂ fluctuation profile in Chamber 3 was created based on the actual data collected from the field. The fluctuation profile varied with canopy growth, so occasional adjustment in CO₂ fluctuation valve was needed for Chamber 3. The air flow in the chambers caused the plants to bend in the early growth stages but became upright with time (Figure 5). For unknown reasons, the PP Systems IRGAs (close path CO₂ analyzers, see CO₂ sensors section) used in chamber 1 & 2 showed some undesirable fluctuations in the CO₂ readings especially at night time, but managed to keep the

median CO₂ value at 610 μmol/mol. Crops grew well inside the chambers (Figure 5) without any heat damage and matured normally.

Conclusion

Chambers had no harmful effects on crop growth and kept the temperature and RH close to the ambient, providing confidence in the chamber design and CO₂ control systems. These chambers with their CO₂ control systems are suitable for testing the biological impacts of fluctuating CO₂ on crops in the field.

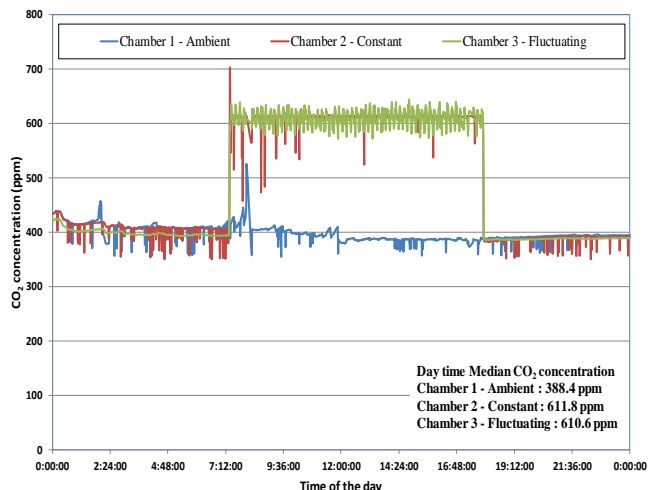


Figure 6. Typical performance of three chambers on 9 August 2015.

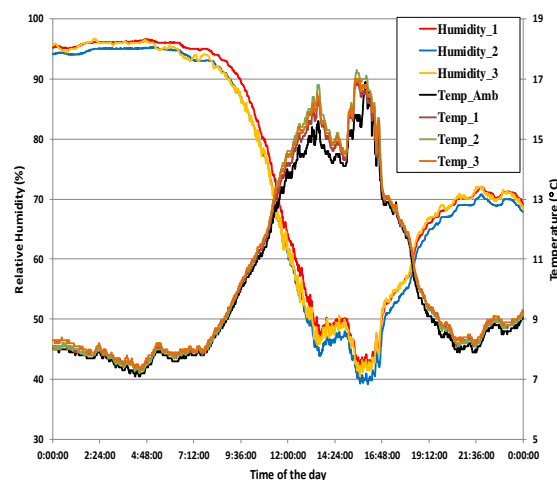


Figure 7. Relative humidity and temperature inside three chambers on 9 August 2015

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