

Soil type influences N₂ fixation in fieldpeas more than elevated CO₂

R D Armstrong¹, M Bourgault² and S.K. Lam³

¹ R D Armstrong Department of Economic Development, Jobs, Transport and Resources, Horsham Victoria 3400, roger.armstrong@ecodev.vic.gov.au

² Faculty of Veterinary and Agricultural Sciences, The University of Melbourne, Creswick VIV 4144.

³ S. K. Lam Faculty of Veterinary and Agricultural Sciences, The University of Melbourne, Richmond VIC 3121.

Little is known about how elevated atmospheric carbon dioxide (eCO₂) affects pulse growth and nitrogen (N) fixation in medium rainfall dryland cropping systems. We examined the growth and N₂ fixation of field-peas (*Pisum sativa* cv. OZ0601) in the SoilFACE array at Horsham (annual rainfall 420 mm). Fieldpeas were grown under either ambient (ca. 390 ppm) or elevated CO₂ (550 ppm) in large (30 cm width x 100 cm long) intact soil cores placed in replicated bunkers in the ground. The intact cores, which maintain the physical and chemical integrity of the soil profile, were collected from 3 soil types: a Calcarosol, a Vertosol and a Chromosol. Soil N concentration of the 3 soils was Chromosol > Vertosol > Calcarosol. The fieldpeas were grown in rotation with wheat (*Triticum aestivum* cv. Yipti) over 2 seasons and N fixation assessed using the natural abundance method. In 2010, the growth and grain yield of fieldpea was increased by eCO₂ and affected by soil type. Neither factor however affected the proportion of plant N derived from N₂ fixation (%Ndfa), although there was a significant interaction between soil type and eCO₂ on the amount of N fixed. In 2011, soil type significantly affected growth, yield and %Ndfa of field pea but eCO₂ had no effect. %Ndfa was strongly related to the amount of NO₃-N in the profile at sowing. N₂ fixation in fieldpeas is, at least in the initial phase of CO₂ treatment, influenced more by soil type (soil N availability) than eCO₂.

Key words

SoilFACE, elevated CO₂, N₂ fixation

Introduction

Future increases in atmospheric CO₂ offer the chance to greatly increase plant productivity via effects on a range of plant physiological processes including photosynthesis and water relations (Kimball *et al.* 2002). These increases however are contingent on adequate supplies of available nitrogen (N) (Newton *et al.* 2007). The productivity of Australian cropping systems is heavily reliant on soil N supply, with a large proportion of this N traditionally coming from biological fixed N including that from grain legumes (Angus 2001, Evans *et al.* 2001). The rate of N₂ fixed, and the value of the N derived from legumes to subsequent crops, can vary markedly with seasonal conditions and site/soil (Kirkegaard *et al.* 2008).

Growth stimulation by elevated CO₂ (eCO₂) is generally greater in legumes than non-legumes, probably as a result of maintaining high leaf N status (Kimball *et al.* 2002). FACE (Free Air Carbon Dioxide Enrichment) studies have revealed many factors operating in the open field situation that cannot be identified in other types of studies (Leakey *et al.* 2009). There have been very few studies however (other than for soybeans) of the response of grain legumes to eCO₂ in the field (Rogers *et al.* 2009). In this paper we report an experiment that examined the effect of eCO₂ on the growth and N₂ fixation of a pulse (field pea) grown in rotation with wheat in three different soil types in the SoilFACE array.

Methods

SoilFACE, located near Horsham (36°45'S, 142°06'E; 127m elevation) is a FACE array based on the use large intact cores (30 cm diameter x 100 cm deep cased in a PVC sleeve), that maintain the physicochemical integrity of the soil profile, to assess interactions between soil type and eCO₂ on crop growth. Cores were collected from the Victorian Mallee (Calcarosol), Wimmera (Vertosol) and High Rainfall Zone (Chromosol) (Table 1) with experimentation commencing in 2009. The cores were placed in 8 bunkers sunk into the ground (the top of the cores are at ground level). Four bunkers are maintained at ambient CO₂ (ca. 390 ppm) whilst four are maintained at 550 ppm atmospheric CO₂ (eCO₂) as per Mollar *et al.* (2009). The experimental design consisted of field pea (cv. OZ00601) grown in rotation with wheat (cv. Yipti) x 3 soil types x eCO₂/ambient CO₂ x 4 reps in a split plot design.

At peak flowering one intact core in each treatment was cut at ground level, the material dried at 70°C, weighed and ground. Total N and ¹⁵N enrichment was determined by isotope ratio mass spectrometry (IRMS)

(continuous flow Isotope Cube (Elementar, Germany) coupled with a continuous flow mass spectrometer (Isoprime, United Kingdom) utilizing Dumas flash combustion. The proportion of N derived from fixation was determined using the natural abundance technique (Peoples *et al.* 1989) using wheat collected from cores of the same soil type as a reference plant. At grain maturity plants were cut at ground level prior to drying, grain separated by threshing and total N content of the ground material determined by Leco™ analyser (St Joseph, MI, USA). A basal application of P (single superphosphate equivalent to 15 kg P/ha) was sown with the seed. Prior to sowing each year, soil was collected at 0-10, 10-20, 20-30, 30-50, 50-70, 80-95 cm using a thin walled tube (42 mm cutting tip) and gravimetric soil water (following drying at 105°C) and NO₃-N measured by extraction (1:10) in 2 M KCl followed by colorimetric analysis (Searle 1984) on a Flow Injection Analyser. Annual and growing season rainfall (April – November: GSR) at the site was 559 and 330 mm in 2010 and 507 and 250 mm in 2011, respectively.

Table 1: Characteristics of 3 soils used in SoilFACE

Soil	pH (CaCl ₂) 0-10 cm	EC(1:5) 80-100 cm (dS/m)	ESP 80-100 cm (%)	Total N 0-10 cm (%)	Total C 0-10 cm (%)
Chromosol	4.5	0.16	4.3	0.40	4.66
Vertosol	7.7	1.85	20.0	0.08	1.10
Calcarosol	5.9	0.53	7.5	0.05	0.64

Results

In 2010 (second season after CO₂ treatment commenced), eCO₂ stimulated the growth at flowering and maturity, early N content and grain yield of field pea (Table 2). Dry matter production and grain yield were strongly affected by soil type in the general order Vertosol > Chromosol > Calcarosol. The proportion of plant N derived from N₂ fixation at flowering was significantly affected by soil type (lowest on the Chromosol and highest on the Vertosol and Calcarosol soils) but CO₂ treatment had no effect. As a consequence, there was a significant interaction between the CO₂ and soil type on the amount of N fixed (kg/ha) at flowering in the eCO₂ Vertosol cores being greater than ambient CO₂ cores, which in turn were higher than that in the Chromosol soil, regardless of CO₂ treatment. In 2011, the relative growth and grain yield of fieldpeas in the Calcarosol cores increased especially compared to the Chromosol cores, to be no different than the Vertosol. CO₂ treatment however had no effect on the growth, yield or rate of N₂ fixation of fieldpeas.

The large differences in background soil N and C at the commencement of SoilFACE (Table 1) were reflected in the amount of soil NO₃-N (0-90 cm) measured at sowing in 2010 with the Chromosol cores having much higher NO₃-N (146 kg/ha) than the Vertosol (64 kg/ha), which in turn was higher than the Calcarosol soil (24 kg/ha) (data not presented). Neither CO₂ nor previous crop had any significant effect on NO₃-N. In 2011, (3rd season of experimentation), this effect on NO₃-N at sowing persisted with Chromosol > Vertosol > Calcarosol and generally no effect of CO₂ or previous crop: the exception was that the large amount of NO₃-N recorded in eCO₂ Chromosol cores following wheat in 2010. These differences in soil NO₃-N were strongly related (R² = 0.73) to the %Ndfa of the field peas (Figure 1).

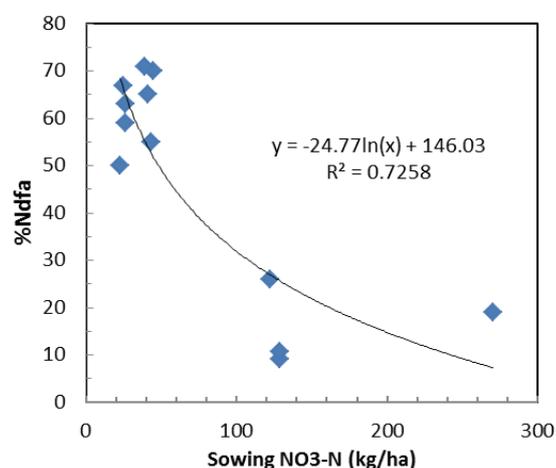


Figure 1: Relationship between profile NO₃-N at sowing and the proportion of legume N derived from N₂ fixation (%Ndfa) of field peas (pooled across years, soil and CO₂ treatments).

Table 2: Influence of CO₂ (ambient and eCO₂) and soil (Calcarosol, Vertosol and Chromosol) on the proportion of N derived from N₂ fixation and the amount of N fixed at flowering and the total dry matter, grain yield and N uptake of field pea at maturity in SoilFACE (2010 and 2011)

CO ₂	Flower DM (g/core)	Flower N Content (g/core)	%Ndfa	N fixed (g/core)	Maturity DM (g/core)	Grain yield (g/core)	Maturity N (mg/core)
2010							
Calcarosol							
Amb	32.6	710	49.9	379	46.9	25.3	nd
eCO ₂	28.1	580	58.9	334	49.7	28.5	nd
Vertosol							
Amb	60.1	1330	55.0	451	75.2	39.5	nd
eCO ₂	96.0	1950	64.6	995	91.0	48.7	nd
Chromosol							
Amb	47.1	1020	9.1	81	51.0	23.4	nd
eCO ₂	57.7	960	10.7	63	66.2	32.6	nd
l.s.d. (5%)	Soil x CO ₂ = 21.1	CO ₂ ns Soil = 339	Soil = 29.7	Soil x CO ₂ = 347	CO ₂ = 7.8 Soil = 9.5	CO ₂ =3.95 Soil = 4.9	nd
2011							
Calcarosol							
Amb	52.9	1112	69.9	780	58.2	30.3	1256
eCO ₂	51.9	1001	71.4	716	73.6	37.8	1334
Vertosol							
Amb	60.6	1388	62.6	858	70.9	35.9	1392
eCO ₂	50.3	1075	67.0	717	80.9	43.7	1802
Chromosol							
Amb	34.0	699	25.6	138	53.3	26.2	973
eCO ₂	26.2	555	18.6	108	60.2	29.2	1068
l.s.d. (5%)	Soil = 14.6	Soil = 307	Soil = 27.2	Soil = 343	Soil = 10.1	Soil = 3.9	Soil = 387

Discussion

Any stimulatory effect of eCO₂ on plant production is strongly related to adequate N supplies (Leakey et al. 2009). This relationship between N supply and eCO₂ will most likely be even more important in the dryland cropping systems of southern Australia as N inputs form a significant proportion of grain grower's variable cost inputs, and N derived from N₂ fixation provides a major proportion of N supplies in many cases. In this study we found that effect of eCO₂ on the dry matter and yield of field peas varied with season, being more important in 2010 when soil water availability was much above long-term average than in 2011.

However eCO₂ had no effect on %Ndfa in either year, although it did increase the amount of N fixed in 2010 as a result of the greater dry matter produced. In contrast, soil type had a much greater effect on growth and yield as well as %Ndfa in both 2010 and 2011. This result is in general agreement with other studies of N dynamics of legumes under eCO₂ that found that whereas the amount of N fixed increases by an average of 38% under eCO₂, the change in %Ndfa is non-significant (Lam *et al.* 2012). Legume N₂ fixation is very sensitive to soil nitrate concentration (Herridge et al. 1998) and in our study we found a strong negative relationship between NO₃-N at sowing and %Ndfa, that appeared to override eCO₂ treatment (Figure 1).

There is a general conclusion that, at least in grasslands, the availability of N for plant growth declines with time under eCO₂ (Progressive Nitrogen Limitation) (Newton *et al.* 2010). Our study found that eCO₂ had little consistent measurable effect on %Ndfa whereas soil type (and especially background N content) had a major impact during the first 3 years after CO₂ treatment commenced. In soils like the Wimmera Vertosol where the background C and N content is relatively low after decades of continuous cropping, even small changes in N₂ fixation rates of pulses may have significant impacts on productivity as the relative value of N derived from legumes to subsequent crops may be reduced in future environments with eCO₂ (Lam et al. 2013).

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