

TOWARDS GENETIC MODIFICATION OF RUMEN BACTERIA FOR IMPROVED PASTURE FIBRE DIGESTION

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Improved microbial fibre digestion holds the potential to increase production in the Australian beef and sheep industries. The introduction of genes for improved fibre-degradation to rumen bacteria has been impeded by the lack of suitable genetic transformation systems. Genetic transformation of *Butyrivibrio fibrisolvens* is prerequisite for the expression of recombinant fibre-degrading genes within this group of rumen microorganisms. *Escherichia coli*/*B. fibrisolvens* shuttle vectors have been described for *B. fibrisolvens* (1, 2, 3), however these vectors are limited in host range or utility. We have investigated the potential of the broad host range plasmid pUB110 to enable the expression of recombinant fibre-degrading genes in *B. fibrisolvens* strains.

MATERIALS AND METHODS

B. fibrisolvens strains were grown anaerobically in M2 medium (4) containing 0.5% w/v cellobiose. Plasmid DNA was isolated by alkaline lysis (5) from *Bacillus subtilis* or transformed *B. fibrisolvens* strains and then used to electroporate *B. fibrisolvens* (2).

RESULTS AND DISCUSSION

B. fibrisolvens strains isolated from the rumen of sheep (AR9, AR11, AR51), arctic reindeer (E14), cattle (H17c), white-tailed deer (OB156), water buffalo (LP1028), and goat (LP1309) were successfully transformed with the plasmid vector pUB110 and/or recombinant derivatives carrying hemicellulase (pUBxynA) or cellulase (pUBcelD) cDNAs isolated from the anaerobic rumen fungus *Neocallimastix patriciarum* (Table 1).

Table 1. *B. fibrisolvens* strains transformed with pUB110 and recombinant derivatives.

Strain(s)	Plasmid Constructs		
	pUB110	pUBxynA	pUBcelD
AR9, E14	+		
AR11	+	+	
AR51, H17c, LP1028, LP1309, OB156	+	+	+

The successful transformation of *B. fibrisolvens* strains with recombinant fibre-degrading genes will allow testing of the hypothesis that modified rumen bacteria may contribute to the improvement of fibre-digestion in livestock.

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